

Short Communication

Plasma choline concentration varies with different dietary levels of vitamins B₆, B₁₂ and folic acid in rats maintained on choline-adequate diets

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Abstract

Choline is an important component of the human diet and is required for the endogenous synthesis of choline-containing phospholipids, acetylcholine and betaine. Choline can also be synthesised *de novo* by the sequential methylation of phosphatidylethanolamine to phosphatidylcholine. Vitamins B₆, B₁₂ and folate can enhance methylation capacity and therefore could influence choline availability not only by increasing endogenous choline synthesis but also by reducing choline utilisation. In the present experiment, we determined whether combined supplementation of these B vitamins affects plasma choline concentration in a rat model of mild B vitamin deficiency which shows moderate increases in plasma homocysteine. To this end, we measured plasma choline and homocysteine concentrations in rats that had consumed a B vitamin-poor diet for 4 weeks after which they were either continued on the B vitamin-poor diet or switched to a B vitamin-enriched diet for another 4 weeks. Both diets contained recommended amounts of choline. Rats receiving the B vitamin-enriched diet showed higher plasma choline and lower plasma homocysteine concentrations as compared to rats that were continued on the B vitamin-poor diet. These data underline the interdependence between dietary B vitamins and plasma choline concentration, possibly via the combined effects of the three B vitamins on methylation capacity.

Key words: B vitamins: Plasma choline: Plasma homocysteine: Methylation capacity: Rats

Choline is an important nutritional component for humans⁽¹⁾ and is required for the endogenous synthesis of phosphatidylcholine (PC), lysophosphatidylcholine, choline plasmalogen and sphingomyelin, which are all essential components of cell membranes. Choline is also a precursor for the biosynthesis of the neurotransmitter acetylcholine and can be metabolised in the liver to betaine, which under certain conditions serves as a source of methyl groups. Choline can be derived not only from the diet, but also from *de novo* synthesis, principally in the liver. Endogenous choline is produced by the sequential methylation of phosphatidylethanolamine (PE) to PC. Choline is then liberated from the

newly formed PC and is released into the bloodstream. This is the only known endogenous pathway for choline biosynthesis in animals⁽²⁾.

The methylation of PE to PC is catalysed by PE-*N*-methyltransferase (PEMT), which requires the methyl donor *S*-adenosylmethionine (SAM). This reaction is influenced not only by the availability of SAM, but is also inhibited by *S*-adenosylhomocysteine (SAH), and the ratio of SAM:SAH (an indicator of methylation capacity) therefore affects the activity of PEMT⁽³⁾. Moreover, SAH is hydrolysed to homocysteine via a reversible reaction: thus excess homocysteine will result in increased SAH, thereby inhibiting PEMT⁽⁴⁾.

Abbreviations: AD, Alzheimer's disease; PC, phosphatidylcholine; PE, phosphatidylethanolamine; PEMT, phosphatidylethanolamine-*N*-methyltransferase; SAH, *S*-adenosylhomocysteine; SAM, *S*-adenosylmethionine.

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Vitamins B₆, B₁₂ and folate can support PEMT activity both by reducing homocysteine levels and by increasing methionine levels, resulting in an increased SAM:SAH ratio, and thus an increased methylation capacity. An alternative pathway for the regeneration of methionine from homocysteine in the liver is provided by the betaine–homocysteine methyltransferase pathway which involves the conversion of choline in the methyl donor betaine. This pathway thus utilises choline⁽²⁾ and is assumed to be less significant for maintaining methylation capacity when endogenous concentrations of B vitamins are sufficient^(5–8). B vitamins may therefore affect choline availability not only by increasing endogenous choline synthesis through the PEMT pathway, but also by reducing choline utilisation by the betaine–homocysteine methyltransferase pathway.

The interdependence between B vitamin intake and choline status is suggested by several animal^(9–16) and human^(7,17–19) studies. However, to date, research has concentrated on the effects of single B vitamins and on conditions of inadequate choline intake, but none of these reports has shown direct effects of B vitamins on plasma choline. The aim of the present experiment was to investigate the interdependence between dietary vitamins B₆, B₁₂ and folate and plasma choline concentration in a dietary background of choline adequacy and mild B vitamin deficiency. The mild B vitamin deficiency was induced with a B vitamin-poor diet which was previously shown in our laboratory to induce a moderate increase in plasma homocysteine concentration (N. van Wijk and L. M. Broersen, unpublished results) and is thought to be relevant for conditions and/or diseases which are associated with mild B vitamin deficiencies and/or moderate increases in plasma homocysteine, such as ageing or Alzheimer's disease (AD)^(20,21). To this end, we measured plasma choline and homocysteine concentrations in rats that had consumed supplemental amounts of vitamins B₆, B₁₂ and folic acid with recommended amounts of dietary choline after a mild B vitamin deficiency was induced.

Materials and methods

Animals

A total of thirty-two male Sprague–Dawley rats (Charles River, Wilmington, MA, USA), aged 6 weeks at arrival, were housed in pairs at room temperature, under 12 h light–12 h dark cycles. Animals had free access to food and water. Body weight and food intake were registered once a week. Experiments were carried out in accordance with the 1996 Guide for the Care and Use of Laboratory Animals (National Institutes of Health) and Massachusetts Institute of Technology policies and were approved by the Committee on Animal Care at Massachusetts Institute of Technology (Cambridge, MA, USA).

Diets

Two different diets with varied vitamins B₆, B₁₂ and folic acid contents were used: (1) B vitamin-poor; (2) B vitamin-enriched. Diets were AIN-93M based⁽²²⁾, isoenergetic and

identical with respect to their protein, carbohydrate, fat, fibre, mineral and choline contents. The vitamin mix (AIN-93-VX) was prepared without vitamins B₆, B₁₂ and folic acid and these vitamins were subsequently supplemented accordingly. Choline was added at AIN-93M levels, i.e. 1.0 g/kg⁽²²⁾, which meets the minimal requirements for rats⁽²³⁾. Diets were formulated with vitamin-free, ethanol-precipitated casein (Harlan Teklad, Madison, WI, USA) and were manufactured by Research Diet Services (Wijk bij Duurstede, The Netherlands; reference no. 1652B).

The B vitamin-poor diet contained low amounts of vitamin B₆ (<0.6 mg/kg), B₁₂ (<1.0 µg/kg) and folic acid (<0.1 mg/kg). No sulfathiazole drugs were added to the diet and therefore a limited amount of folate was still expected to be provided by the gut flora. Induction of vitamin B₁₂ deficiency in the rat is difficult to achieve because of significant endogenous storage of vitamin B₁₂. To attain a moderate reduction of endogenous vitamin B₁₂, the diets were supplemented with 50 g/kg pectin (polygalacturonic acid, high methoxyl, Obipektin[®], NF/USP Citrus; TEFCO FoodIngredients b.v., Bodegraven, The Netherlands), which binds vitamin B₁₂ in the intestine, making it less bioavailable⁽²⁴⁾. Pectin consequently promotes depletion of endogenous vitamin B₁₂ through the enterohepatic circulation of vitamin B₁₂. However, since pectin could affect food intake⁽²⁵⁾, the B vitamin-enriched diet also contained pectin to maintain uniform intakes of the diets. Pectin has minimal effects on vitamin B₁₂ status when the diet contains adequate amounts of this vitamin⁽²⁴⁾.

The B vitamin-enriched diet was supplemented with 20.0 mg/kg vitamin B₆, 0.2 mg/kg vitamin B₁₂ and 4.0 mg/kg folic acid. For each, the diet provided 400% of the recommended daily intake according to the National Research Council report on the nutrient requirements of laboratory animals⁽²³⁾.

B vitamin paucity was first induced in all rats by feeding them the B vitamin-poor diet for 4 weeks. Subsequently, animals were either continued on the B vitamin-poor diet or switched to the B vitamin-enriched diet for another 4 weeks. Previously, it was shown in our laboratory (N. van Wijk and L. M. Broersen, unpublished results) that the B vitamin-poor diet increased plasma homocysteine concentration from 7.3 (SEM 0.4) µM (control levels) up to 10.4 (SEM 0.8) and 9.2 (SEM 0.8) µM after the diet was consumed for 4 and 8 weeks, respectively.

Tissue preparation

After the supplementation period, animals that were fasted for 3–4 h were killed by CO₂ gas inhalation and subsequent decapitation by guillotine. Trunk blood was collected through a funnel into EDTA-containing tubes. After centrifugation at 1750 g for 10 min, plasma was aspirated and analysed for plasma homocysteine and choline.

Plasma-free choline and plasma total homocysteine assay

HPLC-electrochemical detection of plasma-free choline was performed according to a method adapted from Fossati

et al.⁽²⁶⁾. After protein precipitation, samples were centrifuged to remove proteins. The supernatant was injected into the HPLC using a post-column immobilised enzyme reactor, in an on-line enzyme reaction to produce H₂O₂, which was detected electrochemically.

Plasma total homocysteine was determined by fluorometric HPLC as previously described⁽²⁷⁾. Briefly, thiol amino acids (free and protein-bound) were reduced with tri-*n*-butylphosphine. After protein precipitation and centrifugation to remove the proteins, thiol groups were derivatised with 7-fluoro-2-oxa-1,3-diazole-4-sulfonamide reagent. The content of the derivatised thiol amino acids was determined by fluorescence detection with excitation at 385 nm and emission at 515 nm.

Statistical analysis

All statistical analyses were performed using SPSS (version 15.0; SPSS Inc., Chicago, IL, USA). Data were expressed as means with their standard errors. *P*-values <0.05 were considered significant. Variables were checked for normal distribution with Shapiro–Wilk's test. Effects of dietary B vitamins on body weight and food intake were analysed using repeated-measures ANOVA with dietary B vitamins as between-subject factor and week as within-subject factor. Plasma choline and homocysteine concentrations were compared between rats fed the B vitamin-enriched and B vitamin-poor diet using ANOVA. Standard Pearson correlation coefficients were calculated for plasma choline and homocysteine.

Results

At the start of the 4-week supplementation period, animals were randomised into the experimental groups according to their body weights. During the entire experimental period, body weight ($F_{(1,30)} = 0.02$, $P = 0.89$) and food intake ($F_{(1,14)} = 2.92$, $P = 0.11$) were unaffected by dietary B vitamins.

After the supplementation period, analyses of plasma samples revealed that plasma free choline concentration was up to 10% higher in rats fed the B vitamin-enriched diet than in rats that were continued on the B vitamin-poor diet ($F_{(1,30)} = 9.52$, $P < 0.005$; Fig. 1(a)). In addition, plasma total homocysteine concentration was lower in animals receiving the B vitamin-enriched diet as compared to the B vitamin-poor group ($F_{(1,30)} = 56.95$, $P < 0.001$; Fig. 1(b)). A significant negative correlation between plasma concentrations of choline and homocysteine was observed within animals fed the B vitamin-poor diet ($r = -0.545$, $P = 0.029$, inset) but not in the B vitamin-enriched group ($r = 0.006$, $P = 0.98$, inset).

Discussion

The present study investigated the effects of dietary B vitamin supplementation on plasma parameters in a rat model of mild B vitamin deficiency and choline adequacy. The results demonstrate for the first time that both plasma homocysteine and plasma choline concentrations are dependent on dietary intake of vitamins B₆, B₁₂ and folic acid in rats. Rats receiving

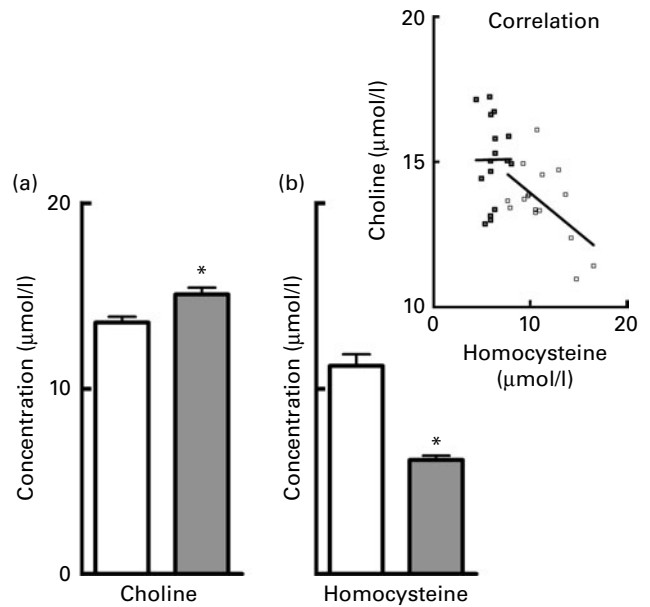


Fig. 1. (a) Plasma-free choline and (b) plasma total homocysteine concentrations and their (inset) correlation in rats that received either a B vitamin-poor diet for 8 weeks (B vitamin-poor, □) or a B vitamin-poor diet for 4 weeks followed by a B vitamin-enriched (■) diet for 4 weeks. Values are means, with their standard errors represented by vertical bars. * Mean values were significantly different ($P < 0.005$).

the B vitamin-enriched diet showed higher plasma choline and lower plasma homocysteine concentrations as compared to rats that were continued on the B vitamin-poor diet. These findings add to previous findings in animals^(9–16) and humans^(7,17–19), and underline an interdependence between dietary B vitamins and plasma choline concentration. A negative correlation between plasma choline and plasma homocysteine was observed solely in the B vitamin-poor group, indicating their mutual dependency on B vitamins. Thus, even at moderately increased plasma homocysteine levels choline status can be compromised, even in a situation where dietary choline intake is considered adequate. This observation is relevant for elderly and AD patients who frequently show mild B vitamin deficiencies and/or moderate plasma homocysteine increases^(20,21) and could therefore be at risk of an affected choline status.

The effects of dietary B vitamin levels on plasma choline concentration may have been mediated by enhancing methylation capacity. An enhanced methylation capacity could influence choline availability not only by increasing endogenous choline synthesis through the PEMT pathway but also by reducing choline utilisation by the betaine–homocysteine methyltransferase pathway. It is important to note that vitamins B₆, B₁₂ and folate all play a crucial role in enhancing methylation capacity. Folate (as 5-methyl-tetrahydrofolate) and vitamin B₁₂ are cofactors in the remethylation reaction catalysed by the enzyme methionine synthase which transforms homocysteine to methionine, from which SAM is subsequently regenerated. Vitamin B₆ is involved in facilitating the reversible conversion of serine to glycine which generates 5,10-methylene-tetrahydrofolate, which can

be reduced to 5-methyl-tetrahydrofolate, i.e. the methyl donor for the remethylation of homocysteine to methionine by methionine synthase. Vitamin B₆ is also the cofactor for the transsulfuration reaction responsible for the irreversible conversion of homocysteine to cysteine, i.e. the clearance of homocysteine. It can be speculated that the effects of the three B vitamins on methylation capacity are additive and therefore have a greater impact on choline metabolism than each B vitamin individually, presumably explaining the observed efficacy in the present experiment encompassing combined supplementation of the three B vitamins.

Poor B vitamin status has been associated with poor cognitive functioning, dementia and AD⁽²¹⁾, while high B vitamin intake has been associated with a reduced risk of AD⁽²⁸⁾. Several hypotheses have been proposed to explain the link between B vitamins and cognitive function, generally invoking effects of B vitamins to reduce presumed neurotoxic homocysteine levels and to increase methylation capacity^(21,29). The present data suggest that this association may at least in part be due to the effects of B vitamins on methylation capacity and subsequent effects on choline availability. Small increases in plasma choline can exert significant effects on brain choline levels⁽³⁰⁾, which in turn control the rates at which it is utilised to form acetylcholine and phospholipids^(31,32).

In the present experiment we showed, for the first time, a direct effect of varying dietary B vitamin levels on plasma choline concentration in rats. The fact that B vitamins influence choline availability might be one of the mechanisms by which B vitamins influence cognition.

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References

1. Zeisel SH, Da Costa KA, Franklin PD, *et al.* (1991) Choline, an essential nutrient for humans. *FASEB J* **5**, 2093–2098.
2. Zeisel SH & Blusztajn JK (1994) Choline and human nutrition. *Annu Rev Nutr* **14**, 269–296.
3. Vance DE, Walkey CJ & Cui Z (1997) Phosphatidylethanolamine *N*-methyltransferase from liver. *Biochim Biophys Acta* **1348**, 142–150.
4. Yi P, Melnyk S, Pogribna M, *et al.* (2000) Increase in plasma homocysteine associated with parallel increases in plasma
5. *S*-adenosylhomocysteine and lymphocyte DNA hypomethylation. *J Biol Chem* **275**, 29318–29323.
5. Park EI & Garrow TA (1999) Interaction between dietary methionine and methyl donor intake on rat liver betaine-homocysteine methyltransferase gene expression and organization of the human gene. *J Biol Chem* **274**, 7816–7824.
6. Holm PI, Bleie O, Ueland PM, *et al.* (2004) Betaine as a determinant of postmethionine load total plasma homocysteine before and after B-vitamin supplementation. *Arterioscler Thromb Vasc Biol* **24**, 301–307.
7. Jacob RA, Jenden DJ, Allman-Farinelli MA, *et al.* (1999) Folate nutrition alters choline status of women and men fed low choline diets. *J Nutr* **129**, 712–717.
8. Yan J, Wang W, Gregory JF, *et al.* (2011) MTHFR C677T genotype influences the isotopic enrichment of one-carbon metabolites in folate-compromised men consuming d9-choline. *Am J Clin Nutr* **93**, 348–355.
9. Troen AM, Chao WH, Crivello NA, *et al.* (2008) Cognitive impairment in folate-deficient rats corresponds to depleted brain phosphatidylcholine and is prevented by dietary methionine without lowering plasma homocysteine. *J Nutr* **138**, 2502–2509.
10. Kim YI, Miller JW, da Costa KA, *et al.* (1994) Severe folate deficiency causes secondary depletion of choline and phosphocholine in rat liver. *J Nutr* **124**, 2197–2203.
11. Akesson B, Fehling C, Jagerstad M, *et al.* (1982) Effect of experimental folate deficiency on lipid metabolism in liver and brain. *Br J Nutr* **47**, 505–520.
12. Chan A, Tchantchou F, Graves V, *et al.* (2008) Dietary and genetic compromise in folate availability reduces acetylcholine, cognitive performance and increases aggression: critical role of *S*-adenosyl methionine. *J Nutr Health Aging* **12**, 252–261.
13. Loo G & Smith JT (1986) Effect of pyridoxine deficiency on phospholipid methylation in rat liver microsomes. *Lipids* **21**, 409–412.
14. She QB, Hayakawa T & Tsuge H (1995) Alteration in the phosphatidylcholine biosynthesis of rat liver microsomes caused by vitamin B₆ deficiency. *Biosci Biotechnol Biochem* **59**, 163–167.
15. Kennedy DG, Blanchflower WJ, Scott JM, *et al.* (1992) Cobalt-vitamin B-12 deficiency decreases methionine synthase activity and phospholipid methylation in sheep. *J Nutr* **122**, 1384–1390.
16. Akesson B, Fehling C & Jagerstad M (1979) Lipid composition and metabolism in liver and brain of vitamin B₁₂-deficient rat sucklings. *Br J Nutr* **41**, 263–274.
17. Abratte CM, Wang W, Li R, *et al.* (2009) Choline status is not a reliable indicator of moderate changes in dietary choline consumption in premenopausal women. *J Nutr Biochem* **20**, 62–69.
18. Abratte CM, Wang W, Li R, *et al.* (2008) Folate intake and the MTHFR C677T genotype influence choline status in young Mexican American women. *J Nutr Biochem* **19**, 158–165.
19. Hung J, Abratte CM, Wang W, *et al.* (2008) Ethnicity and folate influence choline status in young women consuming controlled nutrient intakes. *J Am Coll Nutr* **27**, 253–259.
20. Van Dam F & Van Gool WA (2009) Hyperhomocysteinemia and Alzheimer's disease: a systematic review. *Arch Gerontol Geriatr* **48**, 425–430.
21. Selhub J, Bagley LC, Miller J, *et al.* (2000) B-vitamins, homocysteine, and neurocognitive function in the elderly. *Am J Clin Nutr* **71**, 614S–620S.
22. Reeves PG, Nielsen FH & Fahey GC Jr (1993) AIN-93 purified diets for laboratory rodents: final report of the American Institute of Nutrition *ad hoc* writing committee on the reformulation of the AIN-76A rodent diet. *J Nutr* **123**, 1939–1951.

23. National Research Council (1995) *Nutrient Requirements of Laboratory Animals*, 4th revised ed. Washington, DC: National Academic Press.
24. Cullen RW & Oace SM (1989) Dietary pectin shortens the biologic half-life of vitamin B-12 in rats by increasing fecal and urinary losses. *J Nutr* **119**, 1121–1127.
25. Hove EL & King S (1979) Effects of pectin and cellulose on growth, feed efficiency, and protein utilization, and their contribution to energy requirement and cecal VFA in rats. *J Nutr* **109**, 1274–1278.
26. Fossati T, Colombo M, Castiglioni C, *et al.* (1994) Determination of plasma choline by high-performance liquid chromatography with a postcolumn enzyme reactor and electrochemical detection. *J Chromatogr B Biomed Appl* **656**, 59–64.
27. Krijt J, Vackova M & Kozich V (2001) Measurement of homocysteine and other aminothiols in plasma: advantages of using tris(2-carboxyethyl)phosphine as reductant compared with tri-*n*-butylphosphine. *Clin Chem* **47**, 1821–1828.
28. Luchsinger JA, Tang MX, Miller J, *et al.* (2007) Relation of higher folate intake to lower risk of Alzheimer disease in the elderly. *Arch Neurol* **64**, 86–92.
29. Miller AL (2003) The methionine–homocysteine cycle and its effects on cognitive diseases. *Altern Med Rev* **8**, 7–19.
30. Klein J, Koppen A & Loffelholz K (1990) Small rises in plasma choline reverse the negative arteriovenous difference of brain choline. *J Neurochem* **55**, 1231–1236.
31. Wurtman RJ, Hefti F & Melamed E (1980) Precursor control of neurotransmitter synthesis. *Pharmacol Rev* **32**, 315–335.
32. Wurtman RJ, Cansev M, Sakamoto T, *et al.* (2009) Use of phosphatide precursors to promote synaptogenesis. *Annu Rev Nutr* **29**, 59–87.